

Week 1 (2012.6.11-2012.6.15)

Key Words: [pSB1C3](#)

	Monday 2012.6.11	Tuesday 2012.6.12	Wednesday 2012.6.13	Thursday 2012.6.14	Friday 2012.6.15
Work done		<ul style="list-style-type: none"> . Transformation of the plasmid (pCMV-BMP-2) into E.coli and BBa_J04450-pSB1C3 	<ul style="list-style-type: none"> . Inoculation of BBa_J04450-pSB1C3 - Repeat the transformation of the plasmid (pCMV-BMP-2) 	<ul style="list-style-type: none"> . Inoculation of plasmid (pCMV-BMP-2) - Restreak the plate containing plasmid (pCMV-BMP-2) 	
Result			<ul style="list-style-type: none"> . Got plates with E.Coli carrying BBa_J04450-pSB1C3 (Confirmed by GFP screening) 	<ul style="list-style-type: none"> - Got plates with E.Coli carrying plasmid (pCMV-BMP-2) 	
Discussion					
Remark					

Week 2 (2012.6.18-2012.6.22)

Key Words: [RFP](#), [BMP-2](#), [Pveg](#)

	Monday 2012.6.18	Tuesday 2012.6.19	Wednesday 2012.6.20	Thursday 2012.6.21	Friday 2012.6.22
Work done	<ul style="list-style-type: none"> . Digestion confirmation of plasmid (pCMV-BMP-2) . PCR for front part and back part of BMP-2 from the plasmid (pCMV-BMP-2) (Amplify the BMP-2 from the plasmid, introduce mutation afterwards) 	<ul style="list-style-type: none"> . PCR for P_{veg}-R.B.S. (insert R.B.S. into the biobrick) . Gel check of PCR product P_{veg}-R.B.S. - Gel check for PCR product (BMP-2) 	<ul style="list-style-type: none"> . Gel check of PCR product. P_{veg}-R.B.S. 	<ul style="list-style-type: none"> . gDNA extraction from <i>B. subtilis</i> . Gel check of DNA from genomic extraction - PCR the front part of BMP-2 from the plasmid (pCMV-BMP-2) 	<ul style="list-style-type: none"> . PCR for P_{veg}-R.B.S. (insert R.B.S. into the biobrick) (receive more products) - PCR the back part of BMP-2 from the plasmid (pCMV-BMP-2)) - Digestion of backbone BBa_J04450

					<p>and insert P_{veg}-R.B.S using EcoRI and SpeI.</p> <ul style="list-style-type: none"> Dephosphorylation of PSB1C3 in BBa_J04450
Result	<ul style="list-style-type: none"> The gel photo show the right bands of the plasmid (pCMV-BMP-2) after digestion - 	<ul style="list-style-type: none"> Gel result cannot tell whether PCR P_{veg}-R.B.S is success The gel photo shows the correct band for the front part of BMP-2, but does not show the correct band for the back part of BMP-2 	<ul style="list-style-type: none"> Gel shows PCR product, PCR P_{veg}-R.B.S success. 	<ul style="list-style-type: none"> No DNA found from gDNA extraction 	<ul style="list-style-type: none"> Gel photo does not show the insert P_{veg}-R.B.S and backbone PSB1C3. Save the product
Discussion		<p>No product for the back part of BMP-2. Probably the primer is not correct. In addition, the template may not contain the complementary site as predicted, as the</p>		<ul style="list-style-type: none"> Optimize the protocol for gDNA extraction 	<ul style="list-style-type: none"> Still no solution solving the problem of PCR for back part of BMP-2

		sequence of the template is not documented.			
Remark					

Week 3 (2012.6.25-2012.6.29)					
Key Words: Pveg-R.B.S. gDNA extraction					
	Monday 2012.6.25	Tuesday 2012.6.26	Wednesday 2012.6.27	Thursday 2012.6.28	Friday 2012.6.29
Work done	- gDNA extraction from B. <i>subtilis</i>	. gDNA extraction from B. <i>subtilis</i>	- Ligation of insert P _{veg} -R.B.S and backbone PSB1C3. - Transformation of P _{veg} -R.B.S-PSB1C3 - PCR for signal peptide YdjM from B. <i>subtilis</i> gDNA - Gel check for the PCR YdjM result	- Gradient PCR the whole BMP-2 from plasmid (pCMV-BMP-2) - Gel check for PCR BMP-2 - Check the plate after transformation of P _{veg} -R.B.S-PSB1C3	
Result	. Got gDNA of B.	. Got gDNA of B. <i>subtilis</i>	. Got signal peptide	. No PCR product	

	<i>subtilis</i>		YdjM	from gel photo	
				- Contamination of the transformed plate	
Discussion				- There is some problem on the back part of BMP-2, or the reverse primer	
				- Prepare the plasmid P _{veg} -R.B.S-PSB1C 3 again	
Remark					

Week 4 (2012.7.2-2012.7.6)

Key Words:

	Monday 2012.7.2	Tuesday 2012.7.3	Wednesday 2012.7.4	Thursday 2012.7.5	Friday 2012.7.6
Work done		- Gradient PCR the whole BMP-2 from plasmid	- PCR the whole BMP-2 from BMP-2 extracted in mouse	. PCR YbdN from gDNA of <i>B. subtilis</i> - Gel check for PCR	- Check the plate of transformation P _{veg} -R.B.S-PSB1C

		<p>(pCMV-BMP-2) again</p> <ul style="list-style-type: none"> - Gel check for PCR BMP-2 	<p>gDNA</p> <ul style="list-style-type: none"> - Gel check of PCR product BMP-2 - 	<p>product YbdN</p> <ul style="list-style-type: none"> - Digestion of backbone BBa_J04450 and insert P_{veg}-R.B.S using EcoRI and SpeI. - Dephosphorylation of PSB1C3 in BBa_J04450 - Purification of backbone PSB1C3 and insert P_{veg}-R.B.S - Ligation of P_{veg}-R.B.S and PSB1C3 - Transformation of P_{veg}-R.B.S-PSB1C3 	<p>3</p> <ul style="list-style-type: none"> -
Result	-	- No PCR product	- Whole BMP-2 is amplified from BMP-2 in mouse gDNA	- no product YbdN in the gel photo	- Colonies found in P _{veg} -R.B.S-PSB1C3 3
Discussion				Wrong YbdN primer is added.	

Remark					
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Week 5 (2012.7.9-2012.7.13)

Key Words: P_{veg} -R.B.S-PSB1C3

	Monday 2012.7.9	Tuesday 2012.7.10	Wednesday 2012.7.11	Thursday 2012.7.12	Friday 2012.7.13
Work done	<ul style="list-style-type: none"> . Inoculation 8 colonies from transformed P_{veg}-R.B.S-PSB1C3 - Digestion check for BMP-2 template plasmid (pCMV-BMP-2) using NotI-HF 	<ul style="list-style-type: none"> - Digestion check of P_{veg}-R.B.S-PSB1C3 from 8 colonies using XbaI and NcoI. - Gel check of the digestion product plasmid (pCMV-BMP-2) using NotI-HF 	<ul style="list-style-type: none"> . PCR $YbdN$ from gDNA of <i>B. subtilis</i> - Gel check for PCR product $YbdN$ 		<ul style="list-style-type: none"> - Sequence the plasmid P_{veg}-R.B.S-PSB1C3
Result		<ul style="list-style-type: none"> - Some plasmid P_{veg}-R.B.S-PSB1C3 showed the right band. The cloning success 	<ul style="list-style-type: none"> - Gel showed the right band. Signal peptide $YbdN$ is successfully extracted 		

		- Gel showed the right band of the digested plasmid (pCMV-BMP-2).			
Discussion		Success clone P _{veg} -R.B.S-PSB1C3			
Remark					

Week 6 (2012.7.16-2012.7.20)

Key Words: Overlapping PCR

	Monday 2012.7.16	Tuesday 2012.7.17	Wednesday 2012.7.18	Thursday 2012.7.19	Friday 2012.7.20
Work done	<ul style="list-style-type: none"> Gradient PCR the YdjM-tag-BMP-2 (insert the complementary site of YdjM at the front site of DNA) Gel check of PCR product YdjM-tag-BMP-2 	<ul style="list-style-type: none"> Gel Purify the PCR product YdjM-tag-BMP-2 	<ul style="list-style-type: none"> Gradient PCR the YbdN-tag-BMP-2 (insert the complementary site of YdjM at the front site of DNA) Gel check of PCR product YbdN-tag-BMP-2 Gel Purify the PCR 	<ul style="list-style-type: none"> Overlapping PCR using signal peptide YdjM and DNA YdjM-tag-BMP-2 to make YdjM-BMP-2 Gel check for PCR product YdjM-BMP-2 	<ul style="list-style-type: none"> Digestion of YdjM-BMP-2 and BBa_J04450 using XbaI and PstI Dephosphorylation of PSB1C3 in BBa_J04450 Purification of backbone PSB1C3 and insert

			product YbdN-tag-BMP-2		YdjM-BMP-2 - Ligation of YdjM-BMP-2 and PSB1C3 - Transformation of YdjM-BMP-2-PSB1C3 -
Result	- Gel showed the correct band, PCR success		- Gel showed three bands with one correct.	- Gel showed the right band	- P _{veg} -R.B.S-PSB1C3 has one point mutation on the promoter sequence
Discussion					
Remark					

Week 7 (2012.7.23-2012.7.27)

Key Words:

	Monday 2012.7.23	Tuesday 2012.7.24	Wednesday 2012.7.25	Thursday 2012.7.26	Friday 2012.7.27
Work done	- Check the plate of transformation YdjM-BMP-2-PSB1C3	- Digestion check of the 3 colonies with YdjM-BMP-2-PSB1C3 using StyI	- Inoculation of 8 colonies from transformed YdjM-BMP-2-PSB1C3	- Transformation of the BBa_E1010	- Digestion check of the 8 colonies with YdjM-BMP-2-PSB1C3 using StyI

	- Inoculation of 3 colonies from transformed YdjM-BMP-2-PSB 1C3	- Gel check of the digested product YdjM-BMP-2-PSB 1C3			
Result	- YdjM-BMP-2-PSB 1C3 Transformation success. Colonies found on the plate.	- Gel shows the right band of the digested product.			- Gel shows the right band of the digested product.
Discussion		- Clone YdjM-BMP-2-PSB 1C3 is success. But there is a EcoRI cutting site in the BMP-2 sequence. Wait the mutant BMP-2 to make a standard biobrick			
Remark					

Week 8 (2012.7.30-2012.8.3)

Key Words: YdjM-tag-BMP-2* (*:mutated), YbdN-tag-BMP-2* (*:mutated)

	Monday 2012.7.30	Tuesday 2012.7.31	Wednesday 2012.8.1	Thursday 2012.8.2	Friday 2012.8.3
Work done	<ul style="list-style-type: none"> · Gradient PCR the YdjM-tag-BMP-2* with the template which is mutated BMP-2 - Gel check the PCR product YdjM-tag-BMP-2* - Overlapping PCR YdjM-BMP-2* from signal peptide YdjM and YdjM-tag-BMP-2* - Inoculation of BBa_E1010 		<ul style="list-style-type: none"> - Gradient PCR the YbdN-tag-BMP-2* with the template which is mutated BMP-2 - Gel check the PCR product YbdN-tag-BMP-2* - Overlapping PCR YbdN-BMP-2* from signal peptide YdjM and YbdN-tag-BMP-2* 	<ul style="list-style-type: none"> - Digestion of insert YdjM-BMP-2* and BBa_J04450 using XbaI and PstI - Dephosphorylation of PSB1C3 in BBa_J04450 - Purification of backbone PSB1C3 and insert YdjM-BMP-2* - Ligation of YdjM-BMP-2* and PSB1C3 - Transformation of YdjM-BMP-2*-PSB1C3 	<ul style="list-style-type: none"> - Colony PCR of 3 Colonies from YdjM-BMP-2*-PSB1C3 and 3 colonies from YbdN-BMP-2*-PSB1C3 - Gel check for colony PCR product - Standard assembly of P_{veg}-R.B.S-PSB1C3 to BBa_E1010 - Digestion confirmation of plasmid BBa_B0015 with XhoI

				<ul style="list-style-type: none"> - Digestion of insert YbdN-BMP-2* and BBa_J04450 using XbaI and PstI - Dephosphorylation of PSB1C3 in BBa_J04450 - Purification of backbone PSB1C3 and insert YbdN-BMP-2* - Ligation of YbdN-BMP-2* and PSB1C3 - Transformation of YbdN-BMP-2*-PSB1C3 	
Result	Successfully got the product YdjM-BMP-2* Successfully got the		Successfully get the product YbdN-BMP-2*		- Gel showed the correct band of all YdjM-BMP-2*-PSB1C

	plate BBa_E1010				3 and YbdN-BMP-2*-PSB1C 3 - Confirm the right clone BBa_B0015
Discussion					
Remark					

Week 9 (2012.8.6-2012.8.10)

Key Words: [BBa_E1010](#) (RFP)

	Monday 2012.8.6	Tuesday 2012.8.7	Wednesday 2012.8.8	Thursday 2012.8.9	Friday 2012.8.10
Work done	<ul style="list-style-type: none"> - Ligation of P_{veg}-R.B.S and BBa_E1010 - Transformation of P_{veg}-R.B.S -BBa_E1010 				
Result		- Observe the red			

		colonies with the clone P_{veg} -R.B.S-BBa_E1010. Confirm the function of promoter P_{veg} *(one point mutation) in E.coli			
Discussion					
Remark					

Week 10 (2012.8.13-2012.8.17)

Key Words:

	Monday 2012.8.13	Tuesday 2012.8.14	Wednesday 2012.8.15	Thursday 2012.8.16	Friday 2012.8.17
Work done	- Digestion confirmation of PDG 1661 using EcoRI and BamHI.				
Result					
Discussion					
Remark					

Week 11 (2012.8.20-2012.8.24)

Key Words:

	Monday 2012.8.20	Tuesday 2012.8.21	Wednesday 2012.8.22	Thursday 2012.8.23	Friday 2012.8.24
Work done					
Result					
Discussion					

Week 12 (2012.8.27-2012.8.31)

Key Words:

	Monday 2012.8.27	Tuesday 2012.8.28	Wednesday 2012.8.29	Thursday 2012.8.30	Friday 2012.8.31
Work done					
Result					
Discussion					
Remark					

Week 13 (2012.9.3-2012.9.7)

Key Words:

	Monday 2012.9.3	Tuesday 2012.9.4	Wednesday 2012.9.5	Thursday 2012.9.6	Friday 2012.9.7
Work done					
Result					
Discussion					
Remark					

Week 14 (2012.9.10-2012.9.14)

Key Words: [pTms](#) characterization; Biobrick shipment

	Monday 2012.9.10	Tuesday 2012.9.11	Wednesday 2012.9.12	Thursday 2012.9.13	Friday 2012.9.14
Work done					
Result					
Discussion					
Remark					

Week 15 (2012.9.17-2012.9.21)

Key Words: P_{xyIA} characterization; Biobrick shipment

	Monday 2012.9.17	Tuesday 2012.9.18	Wednesday 2012.9.19	Thursday 2012.9.20	Friday 2012.9.21
Work done					
Result					
Discussion					
Remark					