- Slants for: ——— continued... 6/27/12
  - I 13401 (Amp)
  - I 13401 (Amp)
  - J04450 (Amp)
  - K 174015 (Amp)
- Started Cultures of:
  - K 176012 (Amp)
  - K 299009 (Amp)
  - K 176011 (Amp)
- Poured New Plates (chlor)
- ~~Plasmid Preps for:~~ 6/28/12
  - ~~K 174015 (Amp)~~
  - ~~I 13401 (Amp)~~
  - ~~I 13401 (Amp)~~
  - ~~J04450 (Amp)~~
- Replenished Plasmid Prep Materials

6/28/12
- Plasmid Preps for:
  - K 299009
  - I 13401
  - K 176011
  - K 176012
  - J04450
  - I 13401
  - K ~~299009~~ 174015
- Made liquid cultures for:
  - same 7 as above
- Plates for:
  - same 7 as above
- slants for:
  - same 7 as above
-

6/29/12
- Results
  - Lots of growth for all 7 except...
    - K 174015 (very minimal)
    - leaving it over w/kind
  - J04450 most likely contaminated
    - green fluorescence ...supposed to be red
    - liquid culture/plates discarded.
  - The following showed green fluorescence
    - K 299009
    - K 176012
    - K 176011
  - The following did not:
    - K 174015
    - I 13401

EXP. NUMBER	EXPERIMENT/SUBJECT	DATE
NAME	LAB PARTNER	LOCKER/DESK NO. / COURSE & SECTION NO.

6/28/12
- Ligation from page 44:
  - plates left overnight
  - no visible growth
  - so...
    - repeat transformation
- 3 sets competent cells
- Add 1 mL cold $CaCl_2$ (resuspend)
- incubate on ice 30 min.
- ~~Spin~~ for 30 sec
- resuspend 50λ $CaCl_2$
- ~~flip~~ flick tube
- Add all of ligation DNA
- Incubate ice 30 min.
- Heat shock 42° for 30 sec.
- Chill on ice 5 min.
- Add 1 mL broth to each
- incubate for 1 hour (37°C)
- plate (chlor)

6/29/12
- No growth from transformation cells (either day's -- Procedure 1 or 2)
- Re-ordering Competent cells
-

SIGNATURE	DATE	WITNESS/TA	DATE